

Commentary

What lies between: the evolution of stomatal traits on the road to C₄ photosynthesis

Photosynthesis evolved early in the history of life (Blackenship, 2010), and despite the ubiquity and importance of biological carbon fixation, the process is still far from optimal. The majority of the world's plant species perform C₃ photosynthesis, whereby CO₂ is initially fixed by the enzyme Rubisco (ribulose-1,5-bisphosphate carboxylase/oxygenase). However, Rubisco can also react with O₂, leading to photorespiration, a process which consumes energy and releases previously fixed CO₂. The cost of photorespiration, an inhibition of up to 40% of photosynthesis in today's atmosphere, is thought to have been the driving force behind the evolution of C₄ photosynthesis (Sage, 2004; Gowik & Westhoff, 2011). Species that perform C₄ photosynthesis concentrate CO₂ around Rubisco, thereby greatly enhancing its carboxylation efficiency and largely eliminating photorespiration. This translates into high productivity, and C₄ species constitute some of our most successful crops, including maize (*Zea mays*) and sugarcane (*Saccharum officinarum*), as well our most promising biofuel species, such as switchgrass (*Panicum virgatum*) and *Miscanthus × giganteus*. Because the C₄ photosynthetic pathway has evolved over 60 times in at least 19 families (Sage *et al.*, 2011), the multitude of closely related C₃ and C₄ species provide a powerful tool for understanding the repeated evolution of C₄-associated traits. Many studies focus on comparing characteristics between C₃ and C₄ species in a single lineage, raising the issue of whether traits associated with C₄ species are truly C₄-related or are due to common evolutionary histories or habitat preferences (Edwards & Still, 2008). In this issue of *New Phytologist*, Taylor *et al.* (pp. 387–396) assess differences in stomatal characteristics in a suite of related C₃ and C₄ grasses. The authors show that stomatal traits vary predictably between C₃ and C₄ species, even when phylogeny and growth environment are accounted for in the analysis, thereby clearly attributing differences to functional convergence based on photosynthetic pathway. This work makes a novel contribution to our knowledge of C₄ biology and provides a hitherto missing link between stomatal characteristics and photosynthetic physiology.

In leaves, the uptake of CO₂ is inextricably linked to the loss of water through stomata, with an average of *c.* 2.7 g of carbon fixed per kilogram of water transpired in C₃ plants under non-stressful conditions. Because of this inherent trade-off, the regulation of stomatal conductance can be viewed as an optimization problem, whereby carbon gain per unit water loss is maximized (Cowan & Farquhar, 1977). Accordingly, since C₄

'When do changes in stomatal traits occur as a lineage evolves from an ancestral C₃ state towards full C₄ physiology?'

species have high photosynthetic rates even at low intercellular CO₂ concentrations, they should maintain lower stomatal conductance rates than C₃ species to reduce their transpiration rate and further increase their water-use efficiency (the ratio of photosynthesis to transpiration). Indeed, stomatal conductance rates are reduced in C₄ species (Taylor *et al.*, 2010), and C₄ plants have higher water-use efficiency than C₃ species (Monson, 1989; Sage, 2004; Vogan & Sage, 2011). Consistent with this earlier work, Taylor *et al.* demonstrate that even in a phylogenetically-controlled analysis, C₄ grasses have lower maximum stomatal conductance rates (g_{\max}) than their C₃ relatives. However, the lower g_{\max} of C₄ plants compared to C₃ species might still be due to differences in habitat (Edwards & Still, 2008). Photorespiration is enhanced at high temperatures and low intercellular CO₂ concentrations, and C₄ species tend to grow in hot and arid environments where photorespiratory costs are high (Sage, 2004). Since species in dry environments should restrict water loss regardless of their photosynthetic pathway, Taylor *et al.* also looked at precipitation niche to determine if differences in g_{\max} were explained by water availability. In both mesic and arid environments, the authors found that even when accounting for phylogeny, C₄ species had lower g_{\max} than C₃ species, demonstrating intrinsic differences in stomatal traits between the two groups. Generally, low g_{\max} was achieved in C₄ plants by producing smaller stomata for a given stomatal density, although the authors found differences between lineages, such that some C₄ lines reduced stomatal density, while others preferentially reduced stomatal aperture. Evolutionary convergence towards a common functional solution, but using various anatomical or biochemical means, is a repeated theme in C₄ evolution (Sage, 2004), and this work shows that the same holds true for stomata. These results demonstrate the importance of incorporating both phylogenetic and environmental data into analyses of C₄ trait evolution, as well as filling in a crucial gap in our knowledge of C₄ stomatal characteristics.

When do stomatal characteristics change during C₄ evolution?

While Taylor *et al.* concentrated on C₃ and C₄ monocot species, their work raises interesting questions about the coupled evolution of photosynthetic biochemistry and stomatal characteristics in general. For example, when do changes in stomatal traits, such

| | | Photosynthetic type | | | | |
|-------------------------|---------------------------------------|---------------------|--|---------|----------------------|----------------|
| | | C ₃ | Type I | Type II | C ₄ -like | C ₄ |
| | | | C ₃ -C ₄ Intermediates | | | |
| Carbon-related traits | Enlarged bundle sheath cells | ✗ | ✓ | ✓ | ✓ | ✓ |
| | Photorespiratory CCM | ✗ | ✓ | ✓ | ✓ | ✓ |
| | Increased PEPCase activity | ✗ | ✗ | ✓ | ✓ | ✓ |
| | Active C ₄ cycle | ✗ | ✗ | ✗ | ✓ | ✓ |
| | Mesophyll Rubisco activity | ✓ | ✓ | ✓ | ✓ | ✗ |
| Stomatal-related traits | Increased vein density | ✗ | ✓ | ✓ | ✓✓ | ✓✓ |
| | Increased water-use efficiency | ✗ | ✗ | ✗ | ✓ | ✓ |
| | Stomata regulate lower C _i | ✗ | ✗ | ✗ | ✓ | ✓ |
| | Reduced stomatal density/size | ✗ | ? | ? | ? | ✓ |

Fig. 1 Key traits involved in the evolution of C₄ photosynthesis, with regard to the ancestral C₃ state. Although there is overlap in the categories, traits primarily associated with the development of an optimized C₄ cycle are in blue and traits that emphasize potential changes in stomatal characteristics are in green. While there is no data on when stomatal density or size are altered, changes in other traits related to stomatal regulation between Type II intermediates and C₄-like intermediates imply that stomatal development may also be affected at this transition. PEPCase, phosphoenolpyruvate carboxylase; CCM, carbon-concentrating mechanism; C_i, intercellular CO₂ concentration.

as stomatal density or size, occur as a lineage evolves from an ancestral C₃ state towards full C₄ physiology? In C₄ species, Rubisco is localized to the enlarged bundle sheath cells; phosphoenolpyruvate carboxylase (PEPCase) acts as part of the C₄ cycle which pumps organic acids from the mesophyll into the bundle sheath, where they are decarboxylated (Sage, 2004). In some genera, such as the eudicots *Flaveria* and *Heliotropium*, there are not only C₃ and C₄ species, but also C₃-C₄ intermediate species with varying levels of C₄ anatomy and physiology (Kocacinar *et al.*, 2008; Gowik & Westhoff, 2011; Muhaidat *et al.*, 2011; Vogan & Sage, 2011). Three main groups of intermediates are recognized, each a more derived state than the previous: in Type I intermediates, Rubisco refixes photorespiratory CO₂ in enlarged bundle sheath cells; Type II intermediates also have increased PEP carboxylase activity, indicating some level of a C₄ cycle; and lastly, C₄-like intermediates have a C₄ cycle to concentrate CO₂ in the bundle sheath, but still retain some residual Rubisco activity in the mesophyll (Fig. 1; Sage, 2004; McKown & Dengler, 2007; Kocacinar *et al.*, 2008; Gowik & Westhoff, 2011; Vogan & Sage, 2011).

In lineages such as *Flaveria* where phylogenetic data indicate that C₃-C₄ intermediacy occurs between the ancestral C₃ and the derived C₄ states, intermediates could provide a unique system for studying changes in stomatal development along a gradient of C₄ expression. *Flaveria* has already been used to study stomatal behavior to light and intercellular CO₂ concentrations in C₃, C₃-C₄ intermediates and C₄ species, as well as differences in water-use efficiency (Monson, 1989; Huxman & Monson, 2003; Vogan & Sage, 2011). Water-use efficiency is not enhanced in Type I or II *Flaveria*, or in *Panicum millioides*, a monocot intermediate which lacks a well-developed C₄ cycle (Monson, 1989; Pinto *et al.*, 2011; Vogan & Sage, 2011). As well, C₄ and C₄-like *Flaveria* species regulate stomatal conductance to maintain a lower intercellular CO₂ concentration than more C₃-like intermediates (Vogan & Sage, 2011), implying that changes in stomatal behavior arise late in the evolution of C₄ photosynthesis

(Sage, 2004). Could changes in stomatal anatomy and density also be most pronounced at the transition from Type II intermediacy to a C₄-like state (Fig. 1)? Part of the answer may lie in changes in leaf venation: high vein density appears to be a prerequisite for evolving C₄ photosynthesis in leaves (Sage, 2004), and since stomata generally develop between veins, this may provide a direct mechanism for limiting stomatal density. While C₄-like *Flaveria* species have higher vein density than either Type I or II intermediates (McKown & Dengler, 2007), increases in vein density also occur at the initial transition from C₃ to a Type I intermediate (Sage, 2004; McKown & Dengler, 2007; Gowik & Westhoff, 2011), so venation might constrain stomatal development much earlier in the path towards C₄.

The integration of structure and function has led to exciting insights into C₄ evolution, including links between photosynthetic pathway and hydraulic traits (Kocacinar *et al.*, 2008). The work by Taylor *et al.* highlights that not only are stomata functionally affected by the evolution of C₄ photosynthesis, but that these different functional responses may be underlain by structural changes. Their results also emphasize the need to consider evolutionary history and habitat when attributing traits to convergent evolution. Future research into the generalization of these results in C₄ eudicot lineages with C₃-C₄ intermediates will provide insight on the evolutionary pressures and constraints on stomata and carbon-water trade-offs along the path to C₄.

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